

## SUPPLEMENTARY MATERIAL

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### Chronic cypermethrin exposure alters mouse embryonic stem cell growth kinetics, induces Phase II detoxification response and affects pluripotency and differentiation gene expression

Paola Rebuzzini,<sup>1</sup> Cinzia Civello,<sup>1</sup> Edouard Nantia Akono,<sup>2</sup> Lorenzo Fassina,<sup>3,4</sup>  
Maurizio Zuccotti,<sup>1,4</sup> Silvia Garagna<sup>1,4</sup>

<sup>1</sup>Department of Biology and Biotechnology “Lazzaro Spallanzani”, University of Pavia, Italy

<sup>2</sup>Department of Biochemistry, University of Bamenda, Bambili, Cameroon

<sup>3</sup>Department of Electrical, Computer and Biomedical Engineering, University of Pavia, Italy

<sup>4</sup>Centre for Health Technologies (C.H.T.), University of Pavia, Italy

**Correspondence:** Silvia Garagna, Laboratorio di Biologia dello Sviluppo, Dipartimento di Biologia e Biotecnologie “Lazzaro Spallanzani”, Università degli Studi di Pavia, Via Ferrata 9, 27100 Pavia, Italy. Tel. +39.0382.986323 – Fax: +39.0382.986270. E-mail: [silvia.garagna@unipv.it](mailto:silvia.garagna@unipv.it)

**Key words:** Cypermethrin; embryonic stem cells; cell growth; apoptosis; reactive oxygen species; detoxification response.

**Supplementary Table 1.** Sequences of the forward and reverse primers used for the gene expression analysis.

Gene	Forward	Reverse	Amplicon length - bp
<i>Cat</i>	5' TCATCAGGGATGCCATATTGT 3'	5' ACTCCAGAAGTCCCAGACCAT 3'	101
<i>Gpx1</i>	5' CTACACCGAGATGAACGATCTG 3'	5' CTTGCCATTCTCCTGGTGTC 3'	100
<i>Gpx4</i>	5' TGTGGAAATGGATGAAAGTCC 3'	5' ACGCAGCCGTTCTTATCAAT 3'	100
<i>Sod1</i>	5' TGTCCATTGAAGATCGTGTGA 3'	5' TTGCCCAAGTCATCTGTTTC 3'	94
<i>Sod2</i>	5' CTTACAGATTGCTGCCTGCTC 3'	5' GTAGTAAGCGTGCTCCCACAC 3'	97
<i>Cyp1a1</i>	5' CTGCCTAACTTCCCTGGAT 3'	5' ATGTGCCCTCTCAAATGTC 3'	107
<i>Cyp1b1</i>	5' CCAGGTGCAAACCTTGAGACA 3'	5' TGTCTGCACTAAGGCTGGTG 3'	250
<i>Nqo1</i>	5' TTCTCTGGCGATTCAAGAGT 3'	5' TCTGGTTGTCAGCTGGAATG 3'	306
<i>Gsta1</i>	5' CTTCTGACCCCTTCCCTCT 3'	5' GCCAGTATCTGTGGCTCCAT 3'	183
<i>Ugt1a6</i>	5' ATTGCCTCAGACCTCCTCAA 3'	5' GAGACCATGGATCCAAAGA 3'	236
<i>Oct-4</i>	5' GTGGAGGAAGCCGACAACAATG 3'	5' CACCTCACACGGTTCTCAATGC 3'	107
<i>Nanog</i>	5' CTGCTACTGAGATGCTCGCAC 3'	5' AGCTTTGTTGGACTGGTAG 3'	106
<i>Fgf5</i>	5' TGTGTCTCAGGGGATTGTAGG 3'	5' CATCCGTAATTGGCACTTG 3'	101
<i>Brachury</i>	5' CTCTAAGGAACCACCGGTCA 3'	5' AGCATGGACAGACAAGCAGA 3'	100
<i>Foxa2</i>	5' AAATGAGAGGCTGAGTGGAGA 3'	5' GGCCCACATCTATTAGGGACAC 3'	110
<i>β2m</i>	5' GAATTCACCCCACTGAGACT 3'	5' TGCTTGATCACATGTCTCGAT 3'	103

**Supplementary Table 2.** Distribution of control (CTR) and exposed R1 ESCs and 3T3 cells to cypermethrin (CYP) at their respective LD<sub>50</sub> dose (0.3 or 0.6 mM) during the cell cycle phases.

Samples ( $\mu$ M)	Hours	Frequency of cells (mean $\pm$ SD)			
		G0/G1	S	G2/M	
R1	CTR	12	26.9 $\pm$ 4.4	48.9 $\pm$ 2.8	24.4 $\pm$ 1.6
			29.7 $\pm$ 1.1	47.6 $\pm$ 1.0	22.7 $\pm$ 0.7
	CYP	24	31.6 $\pm$ 2.9	54.2 $\pm$ 3.1	14.2 $\pm$ 0.3
			31.5 $\pm$ 0.5	53.3 $\pm$ 0.7	15.2 $\pm$ 0.3
	CTR	48	29.1 $\pm$ 2.2	52.8 $\pm$ 1.6	18.1 $\pm$ 1.0
			25.7 $\pm$ 0.6*	51.6 $\pm$ 0.3	22.7 $\pm$ 0.6*
	CTR	72	30.9 $\pm$ 1.4	54.0 $\pm$ 1.4	15.1 $\pm$ 0.8
			24.3 $\pm$ 1.3*	54.6 $\pm$ 0.9	21.1 $\pm$ 1.8*
3T3	CTR	8	65.0 $\pm$ 4.0	15.9 $\pm$ 1.9	19.0 $\pm$ 2.7
			62.3 $\pm$ 1.5	17.1 $\pm$ 0.6	20.6 $\pm$ 2.2
	CYP	24	67.1 $\pm$ 1.7	14.3 $\pm$ 1.1	18.6 $\pm$ 1.4
			67.9 $\pm$ 3.2	15.1 $\pm$ 1.4	17.0 $\pm$ 1.8
	CTR	48	68.6 $\pm$ 1.4	14.0 $\pm$ 0.6	17.4 $\pm$ 0.8
			79.2 $\pm$ 1.4*	11.1 $\pm$ 0.8*	9.7 $\pm$ 2.0*
	CTR	72	66.6 $\pm$ 0.7	15.9 $\pm$ 0.4	17.5 $\pm$ 0.9
			68.1 $\pm$ 1.3	15.2 $\pm$ 0.7	16.6 $\pm$ 1.4

\*P<0.001, when compared to CTR.

**Supplementary Table 3.** Fold-change values of redox-related, phase I and phase II gene transcripts, relative to CTR, of R1 cells exposed to 0.3 mM cypermethrin for 12, 24, 48 or 72 h.

Groups	Genes	Hours			
		12	24	48	72
Redox-related	<i>Catalase</i>	1.49 ± 0.64	1.05 ± 0.11	4.06 ± 1.69 <sup>#</sup>	2.67 ± 1.71
	<i>Sod1</i>	1.19 ± 0.47	0.67 ± 0.13*	2.24 ± 0.68 <sup>#</sup>	2.02 ± 0.55*
	<i>Sod2</i>	1.42 ± 0.50	0.98 ± 0.11	3.17 ± 0.70 <sup>#</sup>	1.93 ± 0.77*
	<i>Gpx1</i>	1.29 ± 0.20	0.83 ± 0.10	1.19 ± 0.38	1.52 ± 0.41*
	<i>Gpx4</i>	1.15 ± 0.46	1.10 ± 0.09	2.00 ± 0.01 <sup>#</sup>	1.72 ± 0.20*
Phase I	<i>Cyp1a1</i>	0.95 ± 0.37	0.64 ± 0.28*	0.33 ± 0.32*	0.77 ± 0.30
	<i>Cyp1b1</i>	1.36 ± 0.51	0.64 ± 0.28*	0.47 ± 0.21*	3.52 ± 1.42 <sup>#</sup>
Phase II	<i>Nqo1</i>	0.90 ± 0.09	1.18 ± 0.35	1.22 ± 0.36	2.36 ± 0.31 <sup>#</sup>
	<i>Gsta1</i>	0.94 ± 0.46	1.20 ± 0.40	2.06 ± 1.11 <sup>#</sup>	6.30 ± 2.61 <sup>#</sup>
	<i>Ugt1a6</i>	0.59 ± 0.03*	2.85 ± 0.81 <sup>#</sup>	2.19 ± 0.01 <sup>#</sup>	5.14 ± 2.47 <sup>#</sup>

\*P<0.05; <sup>#</sup>P<0.001.

**Supplementary Table 4.** Fold-change values of redox-related, phase I and phase II gene transcripts relative to CTR samples (set at 1) of 3T3 cell line exposed to 0.6 mM cypermethrin for 8, 24, 48 or 72 h.

Groups	Genes	Hours			
		8	24	48	72
Redox-related	<i>Catalase</i>	0.78 ± 0.29	1.19 ± 0.02*	1.08 ± 0.12	2.03 ± 0.51 <sup>#</sup>
	<i>Sod1</i>	0.94 ± 0.08	0.92 ± 0.08	0.89 ± 0.32	0.94 ± 0.44
	<i>Sod2</i>	0.72 ± 0.16	0.83 ± 0.04*	0.72 ± 0.02*	1.26 ± 0.39
	<i>Gpx1</i>	0.93 ± 0.01	0.63 ± 0.03*	0.76 ± 0.09	0.99 ± 0.14
	<i>Gpx4</i>	0.81 ± 0.07	1.01 ± 0.05	0.86 ± 0.31	1.32 ± 0.12*
Phase I	<i>Cyp1a1</i>	0.91 ± 0.07	1.51 ± 0.25*	1.00 ± 0.01	1.00 ± 0.11
	<i>Cyp1b1</i>	1.00 ± 0.23	0.92 ± 0.07	0.85 ± 0.43	1.15 ± 0.70
Phase II	<i>Nqo1</i>	0.97 ± 0.04	0.98 ± 0.04	1.17 ± 0.22	1.14 ± 0.04
	<i>Gsta1</i>	0.76 ± 0.10	1.14 ± 0.11	3.80 ± 0.04 <sup>#</sup>	2.43 ± 0.42 <sup>#</sup>
	<i>Ugt1a6</i>	1.22 ± 0.21	2.32 ± 0.22 <sup>#</sup>	4.43 ± 1.23 <sup>#</sup>	3.56 ± 1.63 <sup>#</sup>

\*P<0.05; <sup>#</sup>P<0.001.

**Supplementary Table 5.** Fold-change values of pluripotency gene transcripts of R1 ESCs exposed to 0.3 mM cypermethrin for 12, 24, 48 or 72 h relative to control ESCs (set at 1).

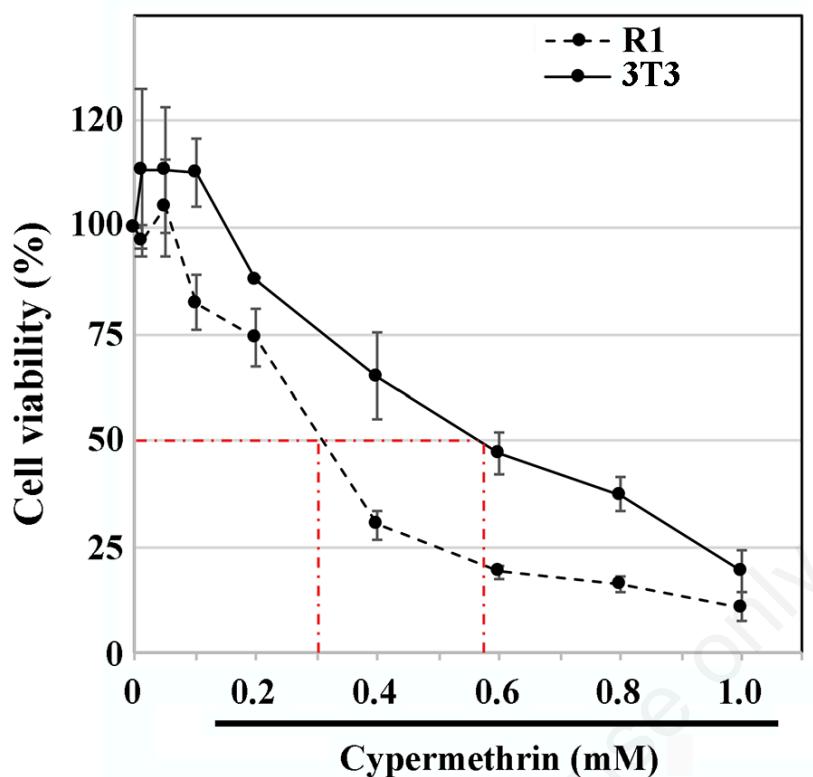
Group	Genes	Hours			
		12	24	48	72
<i>Pluripotency</i>	<i>Oct-4</i>	0.71 ± 0.15*	0.75 ± 0.11*	1.29 ± 0.34*	1.48 ± 0.20*
	<i>Nanog</i>	0.71 ± 0.11*	0.85 ± 0.06*	1.35 ± 0.22*	1.74 ± 0.55*

\*P<0.001.

**Supplementary Table 6.** Fold-change values of early ectoderm, mesoderm and endoderm gene transcripts of EBs differentiated for 5 days from R1 ESCs exposed to 0.3 mM cypermethrin for 12, 24, 48 or 72 h relative to EBs differentiated from control (not exposed) ESCs (set at 1).

Group	Genes	Hours			
		12	24	48	72
<i>Early germ layers</i>	<i>Fgf5</i>	1.00 ± 0.72	0.06 ± 0.01*	0.39 ± 0.08*	0.64 ± 0.34
	<i>Brachyury</i>	0.16 ± 0.05*	1.11 ± 0.04*	1.21 ± 0.26*	0.64 ± 0.06*
	<i>Foxa2</i>	7.28 ± 3.85*	1.82 ± 1.19	7.19 ± 2.69*	3.42 ± 0.12*

\*P<0.001.



**Supplementary Figure 1.**

Dose/response curve to CYP of R1 and 3T3 after 72 h exposure. Data represent the mean  $\pm$  SD of 3 independent experiments.