Immunoreactivities of AR, ERα, ERβ and aromatase in the nuptial pad of Chinese brown frog (Rana dybowskii) during pre-hibernation and the breeding period

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There is a prominent local raised pad called nuptial pad on the forelimb of Chinese brown frog (Rana dybowskii), which is hypothetically concluded as an enhancement of the grip and a spreader of pheromone during the amplexus. In this study, we investigated the immunolocalization and protein expression levels of androgen receptors (AR), estrogen receptor α (ERα), ERβ and aromatase in the nuptial pad of R. dybowskii during pre-hibernation and the breeding period. Histologically, the annual development of the nuptial pad in R. dybowskii is manifested as the larger area of specialized mucous gland and the longer length of papillary epidermal projection during the breeding period. AR, ERα, ERβ and aromatase are present in the stratum granulosum, stratum spinosum, stratum basale and the secretory portion of specialized mucous glands during both periods. Western blotting results confirmed that AR, ERα and ERβ protein levels are higher during pre-hibernation than those during the breeding season. These results suggest that nuptial pad is the direct target organ of androgen and estrogen. Androgen may participate in the regulation of annual development and glandular function of nuptial pad, and estrogen may play an endocrine, autocrine or paracrine role during pre-hibernation and the breeding period.

Key words: Androgen receptor; aromatase; estrogen receptor; nuptial pad; Rana dybowskii.

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Introduction

The nuptial pad is a secondary sexual character of male anurans,\(^1\) and the presence or structural details of nuptial pad are commonly included as standard information in taxonomic descriptions. Nuptial pads are modified areas of skin located at least on the medial margin of finger II of most male anurans, whose invariable characteristic is the presence of sexually dimorphic skin glands (SDSGs) in the dermis, and during the breeding period, nuptial pads develop annually manifesting as thickened epidermis or dermis, or both, when compared with adjacent skin.\(^2\) In the nuptial pads of most species, these SDSGs correspond to specialized mucous glands (SMGs) that consist of a secretory portion, a neck and an intraepidermal duct, which allows molecules synthesized and stored in the SMGs to be channeled to the pad's surface during amplexus and may play an important role in pheromone secretion during courtship.\(^2\) Moreover, further study indicated that amplexus are secreted at papillary epidermal projections (PEPs) of the male nuptial pad which only secrete during the breeding period, the structures of which show similarities with the plectodontid modulating factors (PMF), and the results demonstrate the possible existence of an ancient pheromone system in amphibians.\(^4\) Interestingly, testosterone could deal several regulating effects on the nuptial pad including increases in thickness of both the epidermis and dermis, formation of PEPs, and development of SMGs.\(^5\)

The regulation of androgens on the reproduction of amphibians is manifested in many aspects, including the development of secondary sexual character, the synthesis and delivery of pheromone, the formation of reproductive behavior, etc.\(^5\)\(^,\)\(^,\)\(^,\)\(^,\)\(^6\) Androgen such as testosterone and dihydrotestosterone (DHT) are mainly synthesized by the testis and they are effective in two main ways: directly activate androgen receptors (AR) or conversion into estradiol activate certain estrogen receptors (ER).\(^8\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^9\) Sex steroids are indispensable for the production of male chemical signals related to mating in red-legged salamander. Levels of plasma testosterone are positively associated with development of the male mental gland which secretes courtship pheromones during the breeding season.\(^10\) Testosterone together with prolactin (PRL) can mediate production of the male courtship chemical signal sodefrin in the cloacal gland of male red-bellied newts which has androgen receptors (ARs) and prolactin receptors (PRLRs). Treatment with testosterone and PRL results in the development of the cloacal gland with increased sodefrin mRNA expression and sodefrin content, and testosterone is required for PRL action.\(^11\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^,\)\(^}
Java 1.8.0_172) to calculate the area of every SMG and the length of every PEP.

Immunohistochemistry

After being deparaffinized and rehydrated, the nuptial pad sections were incubated with 10% normal goat serum to reduce background staining caused by the secondary antibody. And then the sections were incubated with primary rabbit anti-human polyclonal antibody (1:500 dilution) against AR (sc-7305) (Santa Cruz Biototechnology, Santa Cruz, CA, USA), ERα (MC-20) (Santa Cruz Biototechnology), ERβ (H-150) (Santa Cruz Biototechnology), and aromatase (H-300) (Santa Cruz Biototechnology) for 12 h at 4°C. The negative control sections were treated with normal rabbit serum instead of the primary antibody. Then the sections were incubated with second antibody, goat anti-rabbit IgG conjugated with biotin and peroxidase with avidin, using a rabbit ExtrAvidin staining kit (Sigma Chemical Co.), followed by visualizing with 30 mg 3,3-diaminobenzidine (Wako, Tokyo, Japan) solution in 150 ml of 0.05 mol Tris-HCl buffer, plus 30 μl H2O2. Finally, the sections for aromatase were counterstained with hematoxylin solution (Merck, Tokyo, Japan). The specificity of aromatase and ER antibodies in this amphibian was previously confirmed.17 The immunostained slides were examined using a photomicroscope (BX51, Olympus, Tokyo, Japan). The immunohistochemistry staining was determined as positive (+), strong positive (+++), very strong positive (+++), and negative (–). Staining that was weak but higher than control was set as positive (+); the highest intensity staining was set as very strong positive (+++); staining intensity between + and +++ was set as strong positive (++).

Western blot

The nuptial pads were diced into small pieces with a clean razor blade respectively and then homogenized in a homogenizer containing 300 μl of 10 mg/ml phenylmethanesulfonyl fluoride (PMSF) stock and incubated on ice for 30 min while maintaining the temperature at 4°C throughout all the procedures. Nuptial pad homogenates were centrifuged at 12,000 × g for 6 min at 4°C. Protein extracts (25 μg) were blotted with an equal volume of 2 x Laemmli sample buffer. Equal amount of each sample was loaded and run on a 12% SDS-PAGE gel at 18 V/cm, and then transferred to nitrocellulose membranes using a wet transblotting apparatus for 20 min (Bio-Rad, Richmond, CA, USA). The membranes were blocked in 3% BSA for an hour at room temperature. Primary incubation of the membranes was carried out using a 1:500 dilution of rabbit anti-human AR, ERα, ERβ and aromatase antibody, which were the same as used in immunohistochemistry for 12 h and the specificity of aromatase and ER antibodies was also confirmed before.17 Secondary incubation of the membranes was then carried out using a 1:1000 dilution of goat anti-rabbit IgG for AR, ERα, ERβ and aromatase, tagged with horseradish peroxidase for an hour. The membrane was then colored with 10 mg 3,3-diaminobenzidine (Wako, Tokyo, Japan) solution in 50 mL phosphate buffer (0.03 M) plus 3μL H2O2. β-actin was used for the endogenous control. The intensities of the bands were quantified using ImageJ software (ImageJ bundled with 64-bit Java 1.8.0_172).

Statistical analysis

The data were expressed as mean ± standard error of the mean (SEM). The normality and homoscedasticity tests were evaluated by D’Agostino & Pearson test and F test, respectively. And statis-
tical significance between differences groups was ensured by two-tailed unpaired Student’s t-test by GraphPad Prism 7. A value of $p<0.05$ was considered indication of statistical significance.

Results

Morphology

During the breeding period, compared with the finger of female frog (Figure 1a), nuptial pads appearing as thickened skins were clearly identified at the base of the Finger I and extended to Fingers II and III (Figure 1b). Figure 1 c,d shows the posture of amplexus in *R. dybowskii* that the male clasps the female about the back and puts its hands closely to the axillar/pectoral area of the female frog. According to our observation, there was no visible wound on the corresponding position of chest (Figure 1e), same with the other *R. dybowskii* we obtained. Figure 2 a,b shows the appearance of the nuptial pads of *R. dybowskii* during the breeding period, and Figure 2 c,d shows the nuptial pads during pre-hibernation, which cover the entire pre-axial part of the thumb in both periods.

Histology

The nuptial pads of *R. dybowskii* consist of several layers of cells, including the stratum corneum (SC), the stratum granulosum (SG), the stratum spinosum (SS), the stratum basale (SB), the stratum spongiosum (SSP) and the stratum compactum (SCO) (Figure 3). One layer of SC, one layer of SG, one to several layers of SS, and one layer of SB form the epidermis from outside to inside, while SSP and SCO form the dermis part. There are PEPs on the nuptial pads, which are formed by an epidermal and dermal evagination (Figure 3 a,b). Melanophores locate in the stratum spinosum of the dermis (Figure 3 a-d). The morphology of SMGs is alveolar, and SMGs are formed by an intraepidermal duct, a neck and a secretory portion, most of which are located in SSP (Figure 3 b,c,e,f). Compared with pre-hibernation (Figure 3 d,e,f), the area of SMG and the length of PEP during the breeding period are significantly larger and longer (Figure 3 a-c), as shown in Figure 4.

Immunohistochemistry

Immunolocalizations for AR, aromatase, ERα and ERβ in the nuptial pads of *R. dybowskii* during pre-hibernation and the breeding period are present in Figure 5. Compared with negative control sections (Figure 5 i,j,k,l), AR is specifically localized in the SG, SS, SB and the secretory portion of SMGs during pre-hibernation and the breeding period, and during pre-hibernation the positive signal is stronger (Figure 5 a,b). Similarly, aromatase, ERα and ERβ are found in the SG, SS, SB and the secretory portion of SMGs during the breeding period, and are also positioned in the SG, SS, SB and the secretory portion of SMGs during pre-hibernation with a stronger positive stain by ERα and ERβ (Figure 5 c-h). The negative control sections (Figure 5 k and l) were stained with hematoxylin, while Figure 5 i and j were not. The staining results obtained from the images are quantified and summarized in Table 1.

Western blot

The protein levels of AR, aromatase, ERα and ERβ in the nup-
tial pad during pre-hibernation and the breeding period were examined by Western blotting, and the results were shown in Figure 6. The positive bands for AR, aromatase, ERα and ERβ were detected at a molecular weight of 110 kDa, 66 kDa, 56 kDa and 55 kDa respectively (Figure 6 a- d). The protein levels of AR, aromatase, ERα and ERβ in the nuptial pad of pre-hibernation are higher than those of the breeding period and the protein levels of AR, ERα and ERβ show significant difference. The quantification results were normalized to the expression level of β-actin.

**Discussion**

In the present study, we reported the immunoreactivities of AR, ERα, ERβ and aromatase in the nuptial pad of *R. dybowskii*. The histological results show that the area of SMG and the length of PEP during the breeding period are significantly larger and longer than those during pre-hibernation. The immunohistochemical data demonstrate that AR, ERα, ERβ and aromatase are present

![Figure 3](image_url)  
**Figure 3.** Histological structure of *R. dybowskii* nuptial pad by hematoxylin-eosin (H&E) during pre-hibernation and the breeding period. a) The nuptial pad during breeding period. b,c) Partial enlarged views of a. d) The nuptial pad during pre-hibernation. e,f) Partial enlarged views of d. SG, stratum granulosum; SS, stratum spinosum; SB, stratum basale; PEP, papillary epidermal projection; SMG, specialized mucous gland; SSP, stratum spongiosum; SCO, stratum compactum; B, the breeding period; P, pre-hibernation. Scale bars: 50 μm.

![Figure 4](image_url)  
**Figure 4.** Measurement result for the area of SMG and the length of PEP during pre-hibernation and the breeding period. Data were shown as the mean ± SEM (n=3, each period). ***p<0.001; ****p<0.0001. B, the breeding season; P, pre-hibernation.

| Table 1. Immunohistochemical localizations of AR, aromatase, ERα and ERβ in the nuptial pad of *R. dybowskii* during pre-hibernation and the breeding period. |
|---|---|---|---|---|---|
| Antibody | SG | SS | SB | SMG | |
| AR | B | P | B | P | B | P | B | P |
| + | ++ | +++ | ++ | +++ | + | ++ | |
| Aromatase | +++ | + | +++ | ++ | +++ | + | ++ | |
| ERα | ++ | ++ | ++ | ++ | ++ | + | +++ | |
| ERβ | + | ++ | ++ | + | +++ | + | ++ | |
| NC | - | - | - | - | - | - | - | - |

*+, negative staining; ++, positive staining; ++++, strong positive staining; ++++, very strong positive staining; SG, stratum granulosum; SS, stratum spinosum; SB, stratum basale; SMG, specialized mucous gland.*

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Figure 5. Immunolocalizations for AR (a,b), aromatase (c,d), ERα (e,f) and ERβ (g,h) in the nuptial pads of *R. dybowskii* during pre-hibernation and the breeding period. Black arrows indicate the positive signal of AR, aromatase, ERα and ERβ in the SG, SS, SB and the secretory portion of SMGs during pre-hibernation and the breeding period. h, i, j, k: Negative control. k and l were stained with hematoxylin. SG, stratum granulosum; SS, stratum spinosum; SB, stratum basale; SMG, specialized mucous gland; NC, negative control; B, the breeding period; P, pre-hibernation. Scale bars: 50 μm.
in the SG, SS, SB and the secretory portion of SMGs during both periods. The protein levels of AR, ERα, ERβ and aromatase are higher during pre-hibernation than those during the breeding period, and the protein levels of AR, ERα and ERβ in two periods show significant difference. These results suggest that androgen may participate in the regulation of annual development and glandular function of nuptial pad, and estrogen may also play an important role via an endocrine, autocrine or paracrine manner during pre-hibernation and the breeding period.

Histologically the nuptial pads of *R. dybowskii* during the breeding season are significantly increased in size, which are manifested as the larger area of SMG and the longer length of PEP, and the results are similar to other researches about the nuptial pad of anurans. For instance, the researches about the annual patterns of nuptial pad in the male Chinese bullfrog (*R. rugulosa*) find that nuptial pad epidermis and mucous glands develop rapidly in January and February, and during the breeding season (March-July), the well-developed nuptial pad is rough, papillate and thick; mucus glands are hypertrophied and lumen secretions are abundant. The researches in *R. ridibunda* also support these results. Luna et al. studied the morphological variation of the nuptial pads in 26 species of *Phyllomedusinae* (Amphibia: Anura: Hylidae), and confirmed the existence of intraepidermal duct in SMG in the nuptial pad of most species, which is consistent with our findings. Furthermore, Epstein et al. found that the nuptial pads of male *R. pipiens* treated with androgen showed strong morphological evidence of activity. For example, the dermal and epidermal layers were thick, the PEPs were developed and the glands exhibited large acini lined with columnar epithelial cells with well-defined cellular boundaries. Moreover, Willaert et al. also found the nuptial pad of *R. temporaria* cause wounds on the female’s chest during amplexus and hypothesized that the secreted molecules can seep directly into the female’s circulatory system. But according to our results, there is no macroscopical wound on the

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**Figure 6.** Western blotting analysis for AR(a), aromatase(b), ERα(c) and ERβ(d) in nuptial pads during pre-hibernation and the breeding period. Data were shown as the mean ± SEM (n = 6, each period, 4 independent experiments). *p<0.05, ***p<0.001, B, the breeding season; P, pre-hibernation.

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breeding period. According to our protein level results that there is autocrine or paracrine manner during pre-hibernation and the breeding period. These results demonstrate that AR, ERα, ERβ and aromatase are present in the SG, SS, SB and the secretory portion of SMGs during both periods. Previous researches indicate that treatment with androgen, or up-regulation of plasma androgen levels, can promote the development of the nuptial pad, such as darkness, SMG size, and other histological characteristics. Our Western blot results show that the protein levels of AR, ERα, ERβ and aromatase are higher during pre-hibernation than those during the breeding period, and the protein levels of AR, ERα and ERβ in two periods show significant difference. Fasano et al. studied the intratissue feedback mechanisms in the regulation of steroid profiles in the R. esculenta, and concluded that testosterone shows high values during early spring (between February and March) and autumn (between October and November), with a rapid decrease thereafter. Considering the time, we obtained R. dybowskii was April and October, our protein level results that AR expressed significantly higher in pre-hibernation than that in the breeding period consistent greatly with previous researches. Moreover, Delgado et al. found that the environmental temperature and photoperiod seasonally play an important role in the regulation of the annual testicular cycle in R. percii, depending on the phase of the annual reproductive cycle. For example, during pre-breeding period, a shortening in photoperiod causes a reduction in testosterone, while low temperature increases testosterone plasma levels during the post-breeding period. Further researches are still needed to explore the mechanism and physiological significance of this regulation in R. dybowskii.

The expression of aromatase is an indication of estrogen production since the conversion of androgens to estrogen by aromatase is the rate-limiting step of estrogen biosynthesis. Most genes have estrogen response elements, and estrogen autocrine/paracrine regulation of local tissue or organ structure and function is common with widespread estrogen receptor. Fasano et al. studied sites of action of local estradiol feedback mechanism in the testis of R. esculenta and concluded that estradiol inhibits androgen synthesis by decreasing the activity of steroidogenic enzymes starting from 17a-hydroxylase. More interestingly, van Wyk et al. assessed the estrogenic contaminants disruption to androgenic control system in male Xenopus laevis which works mainly through anti-androgenic activity, by using the histology of the androgen-dependent SMG in nuptial pad. Besides, in a similar structure, Zhang et al. concluded that estrogen-related genes may be the main factors regulating the seasonal development of the keratinized nuptial spines on upper jaw in Leptobrachium boringii, by assessing the de novo transcriptome using brain, testis and upper jaw skin and comparing gene expression profiles of these tissues between two critical periods of the spine growth cycle. These researches and our results that ERα, ERβ and aromatase share similar immunolocalizations in the nuptial pad of R. dybowskii suggest that estrogens may participate in the regulation of annual development and glandular function of nuptial pad via an endocrine, autocrine or paracrine manner during pre-hibernation and the breeding period. According to our protein level results that there is no difference in the expression level of aromatase in the two periods, we assume there is also no difference in the level of estrogen. The elevated ER protein level improves the sensitivity to estrogen and increases regulation function of estrogen. Another possible explanation is that post-translational mechanisms may be involved in regulating estrogen synthesis in the nuptial pad of R. dybowskii, according to the study in the hypothalamus of the female R. esculenta. In conclusion, the annual development of the nuptial pad in R. dybowskii is manifested as the larger area of SMG and the longer length of PEP during the breeding period. Nuptial pad is the direct target organ of androgens and estrogens. Androgens may regulate the annual development and glandular function of nuptial pad, and estrogens may participate via an endocrine, autocrine or paracrine manner during pre-hibernation and the breeding period. However, the exact function of nuptial pads and PEPs in male is still unclear, neither the origin of the skin modifications. Also, further researches are needed to figure out chemical composition of secretions in nuptial pad SMGs and their roles in reproduction.

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References


