

Benzo[a]pyrene impairs the migratory pattern of human gonadotropin-releasing-hormone-secreting neuroblasts

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ABSTRACT

Benzo[a]pyrene (BaP) is a widespread pollutant that can act as an endocrine disrupting compound (EDC) and interferes with reproductive function. The central regulatory network of the reproductive system is mediated by gonadotropin-releasing hormone (GnRH) neurons, which originate in the olfactory placode and, during ontogenesis, migrate into the hypothalamus. Given the importance of the migratory process for GnRH neuron maturation, we investigated the effect of BaP (10 μ M for 24 h) on GnRH neuroblasts isolated from the human fetal olfactory epithelium (FNCB4). BaP exposure significantly reduced the mRNA level of genes implicated in FNCB4 cell migration and affected their migratory ability. Our findings demonstrate that BaP may interfere with the central neuronal network controlling human reproduction affecting GnRH neuron maturation.

Key words: GnRH neurons; benzo[a]pyrene; EDCs; cell migration; reproductive function; pollution; hypogonadotropic hypogonadism.

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Introduction

The proper establishment of the gonadotropin-releasing hormone (GnRH) system is essential for the regulation of the reproductive competence. During ontogenesis, GnRH-secreting neurons originate from progenitors located in the olfactory placode and migrate along the route of the olfactory nerves to the hypothalamus where they complete their maturation.¹ GnRH neurons remain quiescent until puberty when under permissive signals start to release, in a pulsatile manner, the GnRH peptide regulating sex hormone production and gametogenesis.² Hence, failure of any step of this process may lead to alteration of sex maturation and reproduction.

Research from the past 20 years have documented that some environmental contaminants act as endocrine disruptors interfering with the reproductive system, especially when the exposure occurs during critical developmental periods, such as fetal and perinatal life.^{3,4} Benzo[a]pyrene (BaP) is a widespread persistent organic pollutant⁵ belonging to the endocrine disrupting compounds (EDCs) family.⁶ Thanks to its lipophilic properties, BaP can easily cross the placenta⁷ and acts on fetal development in utero. Increasing evidence, using animal and *in vitro* models, has shown that several EDCs can alter the neuroendocrine control of reproduction,⁸ but the role of BaP in affecting maturation of the central GnRH system in humans remains to be fully explored.

In this study, we examined the effects of BaP on the survival and maturation of human fetal GnRH secreting neuroblasts representative of developing neurons (FNCB4).⁹⁻¹¹

Materials and Methods

Cell culture and reagents

FNCB4 was previously established, cloned, and propagated *in vitro* from human fetal olfactory epithelium,⁹ and characterized as GnRH secreting neuroblasts.⁹⁻¹¹ Cells, cryogenically preserved, were cultured at 37°C in 5% CO₂ atmosphere in Coon's modified Ham F-12 medium (Sigma-Aldrich Corp., St. Louis, MO, USA) supplemented with 10% fetal bovine serum (Hyclone, Logan, UT, USA). BaP was dissolved in dimethylsulfoxide (DMSO; both Sigma-Aldrich) and used at 10 μM as most commonly chosen concentration for *in vitro* toxicity studies.^{12,13} Endothelin 1 (ET1) was obtained from NovaBiochem (Laufelfingen, Switzerland). Cell viability was determined by MTT assay (Sigma-Aldrich), as previously described.¹⁴ The immunocytochemistry was performed as previously described,¹⁵ using rabbit anti-GnRH1 polyclonal antibody (1:100; Abcam, Cambridge, UK) and Alexa Fluor 488-conjugated as secondary antibody (1:200; Molecular Probes, Eugene, OR, USA). AlexaFluor 488-conjugated phalloidin (Molecular Probes) was used for cytoskeletal F-actin detection. Nuclei were counterstained with DAPI (Molecular Probes). Images were collected using a Nikon Microphot-FXA microscope (Nikon, Tokyo, Japan). Cells with a motile phenotype were identified by the presence of cytoskeletal remodeling including elongated cell bodies, filopodia and lamellipodia and were expressed as percentage of total cells, as counted in ten fields from two separate experiments.

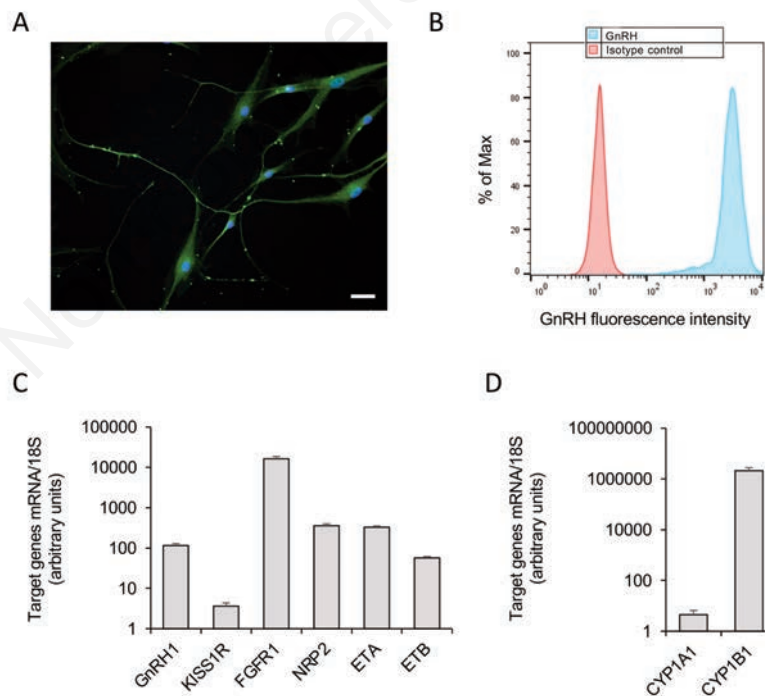


Figure 1. Phenotypic characterization and gene expression profile of FNCB4 cells. A) Representative image of cells expressing GnRH (green) as evaluated by immunofluorescence analysis; DAPI counterstained nuclei; scale bar: 50 μm. B) Representative overlaid histogram of GnRH protein expression (light blue peak) with its isotype control (negative control; pink peak), as detected in FNCB4 cells by flow cytometric analysis. (C-D) Relative mRNA expression by qRT-PCR analysis of target genes normalized over 18S ribosomal RNA subunit, taken as the housekeeping gene, and reported as mean ± SEM (n=9).

Flow cytometry

As already reported,¹⁶ after fixation and permeabilization, cells were incubated with anti-GnRH1 (1:100; Abcam) primary antibody. Alexa Fluor 488 goat anti-rabbit IgG (1:200; Molecular Probes) was used as secondary antibody. Stained cells were analyzed on a FACSCanto II flow cytometer (Becton-Dickinson, San Jose, CA, USA) and analyzed with FloJo software (Ashland, OR, USA).

Quantitative real time RT-PCR (qRT-PCR)

Isolation of total RNA and cDNA synthesis were performed using the RNeasy kit (Qiagen, Hilden, Germany) and the iScript™ cDNA Synthesis Kit (Bio-Rad Laboratories, Hercules, CA, USA) according to the manufacturers' instructions. qRT-PCR was performed as previously described,¹⁷ using a CFX96 Two-Color Real-Time PCR Detection System (Bio-Rad Laboratories). Primers and probes for the target genes were obtained from Life Technologies

(Carlsbad, CA, USA). The 18S ribosomal RNA subunit was used as the housekeeping gene for relative quantitation of the target genes based on the comparative threshold cycle method.¹⁸

Migration assay

Cell migration was assessed with a Boyden chamber system (Greiner BioOne, Essen, Germany) with PET translucent 8 μm pore size membrane. Migrated cells were fixed, stained with 10% Giemsa solution in PBS and counted (3 fields for each insert) in blind under an optical microscope (Carl Zeiss S.p.A., Milan, Italy).

Statistical analysis

Data are expressed as mean ± standard error of mean (SEM). Student's unpaired *t*-tests or one-way ANOVA followed by Tukey's *post-hoc* analyses for multiple comparison were performed, as appropriate, to determine statistical significance ($p < 0.05$), by the Statistical Package for the Social Sciences (SPSS Inc., Chicago, IL, USA).

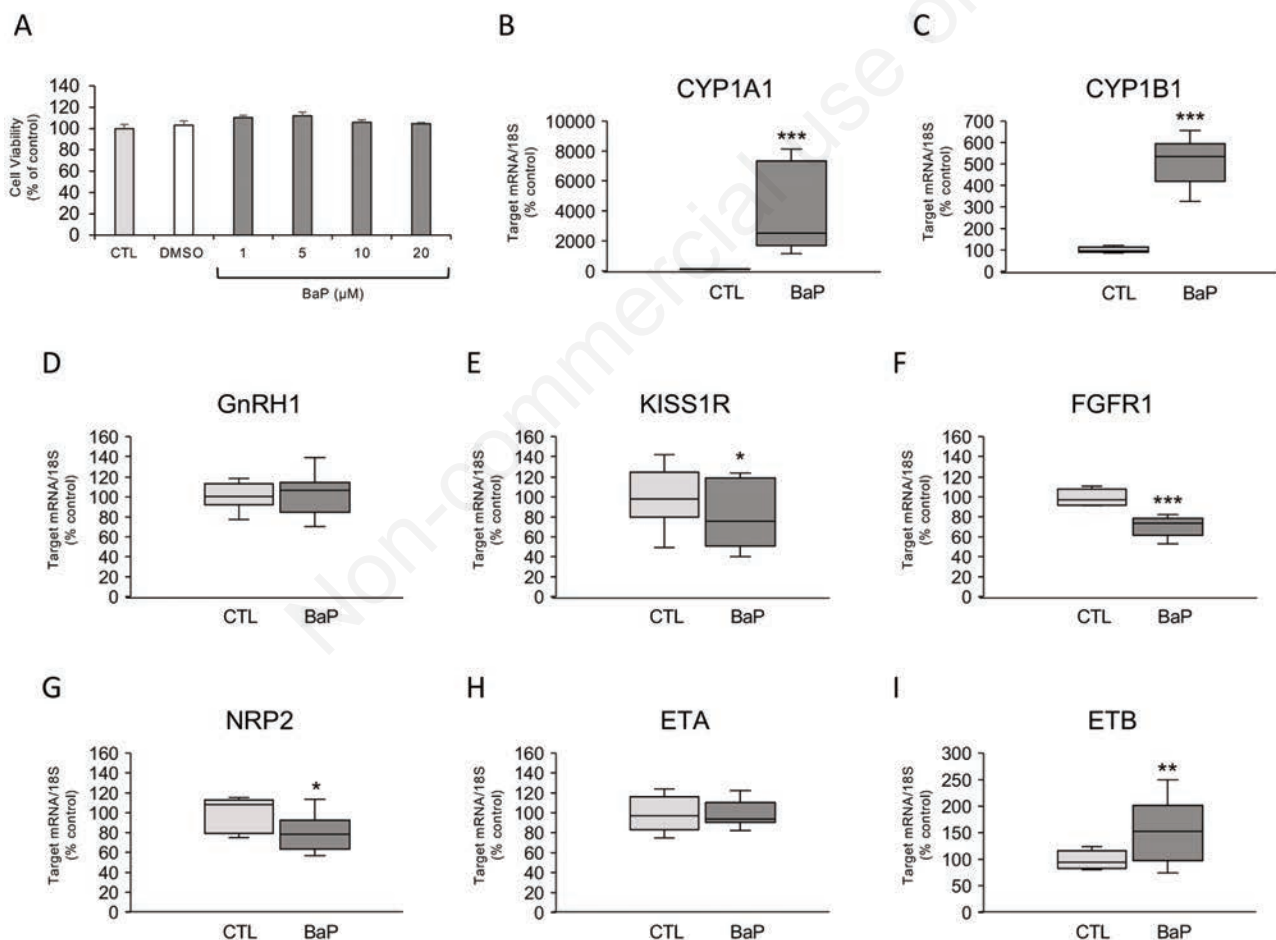


Figure 2. Benzo(a)pyrene effect on FNCB4 cells. A) MTT analysis of FNCB4 cells treated with different concentrations of BaP (1, 5, 10, 20 μM) for 24 h (dark grey bars); cell viability was expressed as percentage of viable cells over control (untreated cells; CTL; light grey bar), taken as 100% (mean ± SEM); vehicle control (0.04% DMSO; white bar) was also performed (n=4). B-I) mRNA expression of target genes in untreated (CTL, light grey box) and BaP-treated (10 μM for 24 h; dark grey box) FNCB4 cells, as detected by qRT-PCR. Data are normalized over 18S ribosomal RNA subunit, taken as the housekeeping gene, reported as percentage of CTL and displayed as mean ± SEM. Statistical analysis was performed using unpaired Student's *t*-test; * $p < 0.05$; ** $p < 0.01$; *** $p < 0.001$; n=12 for CYP1A1 and CYP1B1, n=9 for other genes.

Results

BaP-induced metabolism in FNCB4

The identity of FNCB4 as GnRH neuroblasts was confirmed, as already published.^{11,19,20} Cells showed a strong GnRH immunopositivity (Figure 1 A,B) and the mRNA expression of genes characterizing the GnRH neuroblast phenotype, such as GnRH, KISS1R and genes implicated in GnRH neuron migration (FGFR1, NRP2, ETA and ETB; Figure 1C).

We also verified that FNCB4 expressed CYP1A1 and CYP1B1 mRNAs, the main cytochrome P450 isoforms responsible for metabolic activation of BaP, with CYP1B1 being the most abundant (Figure 1D).

Effect of BaP on FNCB4 phenotype

As detected by MTT assay, 24 h exposure to different concentrations of BaP (1, 5, 10 and 20 μM) did not affect FNCB4 cell viability (Figure 2A) and based on these data the subsequent experiments were performed using 10 μM BaP. Exposing FNCB4 to BaP

(10 μM for 24 h) caused a significant increase in the mRNA level of both CYP1A1 and CYP1B1 (Figure 2 B,C, respectively), as expected.²¹

Concerning the effects on FNCB4 phenotype and function, no significant changes were observed for GnRH1 expression (Figure 2D), whereas KISS1R mRNA was significantly reduced by BaP exposure (Figure 2E). Interestingly, BaP significantly reduced the mRNA expression of FGFR1 and NRP2 (Figure 2 F,G), while it significantly increased ETB mRNA, as compared to untreated cells (Figure 2 H,I).

Effect of BaP on FNCB4 cell migration

To better investigate the impact of BaP on FNCB4 migration, cells were exposed to ET1 (100 nM, 6 h). ET1 significantly increased the percentage of migrated cells when compared to untreated cells, whereas BaP exposure (10 μM , 24 h) significantly prevented this effect (Figure 3A). The evaluation of cellular morphology and cytoskeleton rearrangements indicative of cell movement, confirmed these data. Representative images for immunodetection of F-actin are reported in Figure 3B. Unstimulated cells

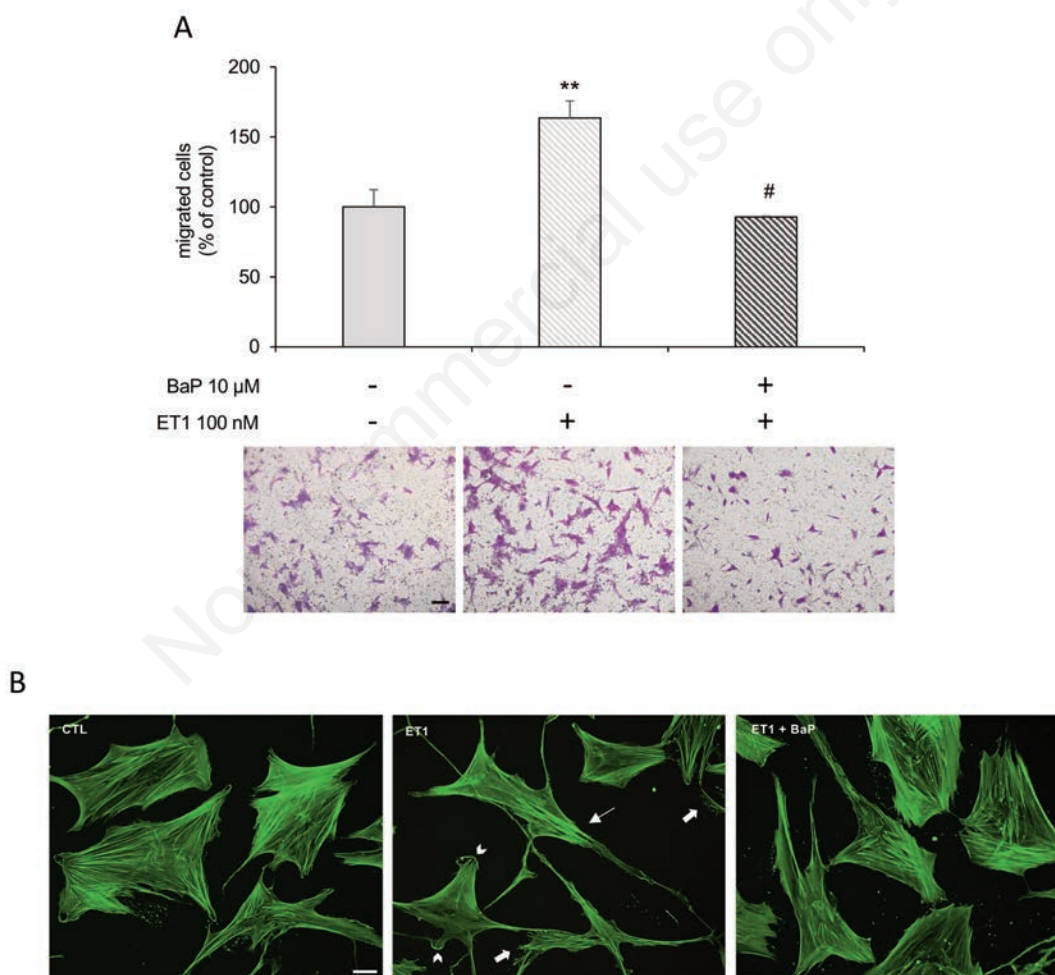


Figure 3. Benzo[a]pyrene alters FNCB4 cell migration. A) Bar graph and representative images showing quantitative analysis of transwell migration assay in FNCB4 cells pre-incubated in presence or absence of BaP (10 μM , 24 h) in serum-free medium, and induced to migration with ET1 (100 nM, 6 h); migrated cells were counted in 3 different fields for each condition, expressed as percentage of migrated cells over control (untreated cells), taken as 100% (mean \pm SEM; scale bar 50 μm); statistical analysis was performed using one-way ANOVA ($p=0.006$) followed by Tukey's post-hoc analysis (** $p<0.01$ vs untreated cells, # $p<0.01$ vs ET1; $n=9$). B) Representative images of cytoskeleton organization of FNCB4 cells exposed to ET1 (100 nM, 24 h) in presence or absence of BaP (10 μM , 24 h) and immunolabeled for F-actin (green) for the detection of microfilaments; arrows, thin arrows, and arrowheads point at cells showing a motile phenotype with elongated cell body, filopodia and lamellipodia, respectively; scale bar: 50 μm .

showed a prevalent actin stress fiber network with scarce actin cytoskeletal remodeling detectable in a low percentage of cells (16.8±4.1%). In presence of ET1 (100 nM, 24h), most FNCB4 cells exhibited various actin-based cell deformations compatible with a motile phenotype, including elongated cell bodies, filopodia and lamellipodia (73.6±3.4%; $p < 0.001$ vs control). The addition of BaP in co-treatment with ET1 for 24 h significantly reduced the percentage of motile cells (23.9±3.1%; $p < 0.001$ vs ET1; Figure 3B).

Discussion

Impaired GnRH neuron migration due to genetic defects is one of the main causes of severe forms of hypogonadotropic hypogonadism (HH) and infertility. Here, we describe for the first time that the migratory properties of human GnRH neuroblasts may be affected by the exposure to BaP, a widespread environmental pollutant, implicated as EDC in altering gonadal function and consequent fertility.⁶ Our results are consistent with previous findings observed in the mouse exposed to other EDCs, such as bisphenol A.^{22,23} Since the cellular model we used is representative of the early GnRH neuron developmental steps,^{9,11,20} our results strongly suggest that BaP exposure may interfere with the correct maturation of the hypothalamic neuronal network controlling the reproductive axis. Moreover, FNCB4 are able to metabolize BaP, as they express CYP1A1 and CYP1B1 enzymes, which convert BaP into bioactive forms.²⁴ The strong induction of both isoforms after BaP exposure indicates that FNCB4 cells may be a direct target of the EDC, which therefore affects the gonadal function not only peripherally but also at the brain level. Indeed, a constitutive expression of CYP1A1 has been reported for neuronal and olfactory bulb cells,^{25,26} as well as CYP1B1 is highly expressed in extrahepatic organs²⁷ including fetal brain.²⁸ Concerning the impact of BaP on FNCB4 phenotype and function, we found negative effects on the expression of cell migration-related genes, such as FGFR1, NRP2 and ETB, with no GnRH mRNA changes. It is well known that FGFR1 mutations cause genetic disorders associated to HH with GnRH deficiency and anosmia indicative of GnRH neuron maturation defects.²⁹ Moreover, the lack of NRP2 is linked to alteration of the normal migratory process of GnRH neurons.³⁰ Accordingly, we demonstrated that BaP exposure significantly reduced both FGFR1 and NRP2 mRNA, as well as the migratory ability. In contrast, BaP significantly induced ETB expression, which we previously demonstrated as being involved in ET1-induced migration in FNCB4 cells.²⁰ ETB induction may be explained by the known ETB-mediated ET1 neuroprotective effects described in the nervous system.^{31,32}

Another interesting finding concerns the observation that BaP exposure significantly reduced KISS1R mRNA. The kisspeptin/KISS1R system is crucially required as master regulator of GnRH release by mature GnRH neurons within the hypothalamus.³³ Hence, we can speculate that during the early steps of fetal development this system is mainly implicated in cell migration mechanisms, which may be more sensitive to BaP, rather than in those regulating GnRH expression. Indeed, kisspeptin has anti-metastatic activity³⁴ and inhibits ET1-induced FNCB4 migration.³⁵ In this context, kisspeptin/KISS1R system could be required for stop signal processes when the neurons reach their destination within the hypothalamus. By reducing KISS1R expression BaP could also interfere with this important step of GnRH neuron maturation.

In conclusion, our findings contribute to identify a novel scenario by which BaP exposure may directly induce neuronal injury in the immature brain interfering with the correct maturation of the hypothalamic network regulating reproduction.

Abbreviations

Benzo[a]pyrene (BaP); endocrine-disrupting compounds (EDCs); endothelin 1 (ET1); gonadotropin-releasing hormone 1 (GnRH1); kisspeptin receptor 1 (KISS1R); fibroblast growth factor receptor 1 (FGFR1); neuropilin 2 (NRP2); endothelin type A receptor (ETA); endothelin type B receptor (ETB); cytochrome P450 1A1 (CYP1A1); cytochrome P450 1B1 (CYP1B1); hypogonadotropic hypogonadism (HH).

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