

Establishment and biological characteristics of a Jingning chicken embryonic fibroblast bank

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Abstract

Using tissue explantation and cryopreservation biotechniques, a Jingning chicken embryonic fibroblast bank was successfully established, which includes 43 embryo samples and a stock of 178 cryovials, each one containing 3.0×10^6 cells. Most of the cells were apparently fibroblasts in their morphology, and the population doubling time (PDT) was about 48 h. The total chromosome number of a diploid cell was 78. According to karyotyping and G-banding, the diploid rate in the cell bank was $97.62 \pm 2.12\%$. The cells were tested for microbial contamination and found free of infections from bacteria, fungi, viruses and mycoplasmas. There was no cross-contamination from other cell lines as revealed by lactate dehydrogenase (LDH) and malate dehydrogenase (MDH) isoenzyme polymorphisms. Six fluorescent proteins were transfected into the Jingning chicken embryonic fibroblasts, and the transfection efficiencies of these genes were between 10.1 and 41.9%. All the tests showed that the quality of the cell line conforms to the quality criteria of the ATCC (American type culture collection). This work succeeded not only in preserving the genetic resources of Jingning chicken, but it also established a new protocol to preserve endangered animal breeds.

Introduction

Genetic diversity of China livestock and poultry had been seriously reduced, and some high-quality species of livestock and poultry are on the brink of extinction, especially due to the exacerbation of environmental pollution and the development of animal husbandry. If these genetic resources are not preserved in any forms before their extinction, they will be irreparably lost; it is therefore crucial to under-

take realistic measures to conserve endangered species.¹ Currently, semen, embryos, genomic and cDNA libraries are all conventional methods to preserve single animals. In addition, since the entire genome of a species is contained in the nucleus of somatic cells, modern cloning techniques have made fibroblasts an attractive resource for safeguarding animal genetic materials.² In addition, the establishment of an animal somatic cell bank can not only preserve this genetic resource at the cell level, but also provide a precious experimental material for investigation in cell biology, genomics and post-genomics, and for embryonic engineering. Jingning chicken, also known as Sanhuang chicken or White-ear chicken, originates in the provinces of Yushan, Shangrao and Guangfeng, China and is characterized by small body, high fecundity and tender flesh. As one of the chicken local breeds in China, Jingning chicken has enjoyed a high reputation in Jiangxi Province ever since the Song dynasty, and was well sold in international market during the Qing dynasty.³ Jingning chicken has been included in the 138 national protected species list by the Chinese Government in 2006.

In the present research, we succeeded in constructing a fibroblast bank through primary explantation and programmed cryopreservation, and tested the quality of the cell lines obtained. The genetic resource of this valuable local breed has thus been preserved through somatic cells for long-term storage. In addition, this technical platform will provide a technical and theoretical support to conserve other animal genetic resources at the cell level.

Materials and Methods

The 8-day old embryos of Jingning chicken used in this research were provided by the chicken breeding farm of the Chinese Academy of Agricultural Sciences, Beijing, China. Except otherwise specifically mentioned, all reagents were obtained from Sigma Chemical Co. (St. Louis, MO, USA).

Isolation and culture of Jingning chicken embryonic fibroblasts

Jingning chicken eggs incubated for 8 days were sterilized using alcohol swabs, and then the embryos were isolated and washed three times with phosphate buffered saline (PBS). Embryos were chopped into pieces of 1 mm³ and placed onto the surface of a tissue culture flask, at 37°C in a humidified air atmosphere containing 5% of CO₂, for 3-4 h. Modified Eagle's medium (MEM) (Gibco, Carlsbad, CA, USA) containing 10% fetal bovine serum

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(HyClone, Logan, UT, USA) was added into the inverted flask and cultured overnight. The medium was refreshed after 2-3 d. The cells were harvested at 80-90% confluence using 0.25% trypsin (m/v) solution and were separated into culture flasks at the ratio of 1:2 or 1:3.^{4,5}

After three passages, the cultured cells were suspended in media containing 40% MEM, 50% fetal bovine serum and 10% dimethyl sulfoxide (DMSO). Cells in logarithmic growth phase were counted with a hemocytometer and adjusted to a density of $3-4 \times 10^6$ viable cells per mL. Aliquots of the cell suspension were transferred into sterile cryovials labeled with the breed name, gender, freezing serial number and date. The vials were sealed and kept at 4°C for 20-30 min to equilibrate DMSO, put into a programmed preservation system, and finally transferred to liquid nitrogen for long-term storage.⁶ Vials taken from the liquid nitrogen were thawed in a 42°C water bath, then transferred to flasks with MEM containing 10% fetal bovine serum and cultured at 37°C with 5% CO₂. The medium was renewed after 24 h.

Estimation of cell viability by trypan blue exclusion test

Viabilities before freezing and after thawing were determined using the Trypan blue exclusion test. The number of non-viable cells was determined by counting 1000 cells.⁷

Growth dynamics

Cells were plated onto 24-well microplates (Corning, Santa Clara, CA, USA) at the density of approximately 1.5×10^5 cells per well, cultured for 7 d and counted every day (3 wells each time) afterwards. The mean cell counts at each time point were then used to plot a growth curve, based on which the population doubling time (PDT) was calculated.⁷⁻⁹

Microbial analysis

Detection of bacteria and fungi

The cells were cultured in complete MEM (Gibco, Carlsbad, CA, USA) media free of antibiotics and observed for the presence of bacteria and fungi at 3 d after subculture, according to the method described by Doyle *et al.*¹⁰

Detection of mycoplasmas

According to the protocol of the American type culture collection (ATCC), the cells were cultured in antibiotic-free medium for at least 1 week, and then fixed and stained with Hoechst 33258 (Invitrogen, Carlsbad, CA, USA).¹¹

Virus detection

Hay's hemadsorption protocol was used to examine the samples for cytopathogenesis using phase-contrast microscopy (Olympus Corp., Tokyo, Japan).^{12,13}

Karyotyping and chromosome analysis

Chromosome spreads were prepared, fixed and stained following standard methods.¹⁴ The cells were incubated, when 70-80% confluent, with $0.1 \mu\text{g/mL}$ of colchicine for 4 h. Hypotonic treatment is a major influential factors in karyotyping, which aims at swelling the cells and spreading the chromosomes; to obtain good metaphase spreadings, the time of the hypotonic treatment was tightly controlled within 40 min. Cells were harvested when 80-90% confluent, subjected to hypotonic treatment and fixed, and the chromosome numbers were counted from 100 spreads after Giemsa staining, using an $100\times$ oil immersion objective. Relative length, arm ratio and centromeric index, and kinetochore type were calculated according to the protocol described by Kawarai *et al.*¹⁵

Isoenzyme polymorphisms

Isoenzyme patterns of lactic dehydrogenase (LDH) and malic dehydrogenase (MDH) were detected using vertical slab non-continuous polyacrylamide gel electrophoresis (PAGE) assay. According to the method described by Simpson,¹¹ the cells were harvested, and protein extraction solution (0.9% Triton X-100,

0.06 mM NaCl:EDTA in mass ratio 1:15) was added after the cell concentration was adjusted to 5×10^7 cells/mL. Then the mixture was centrifuged and the supernatant was aliquoted and stored at -80°C . Liquid sucrose [40% (m/v)] and the samples were mixed [1:1 (v/v)] and then loaded into the individual lanes of the polyacrylamide gel.¹¹

Expression of fluorescent proteins in Jingning chicken fibroblasts

According to the methods described by Wu *et al.*,¹³ three fluorescent proteins (pDsRed1-N1, pEGFP-N3 and pEYFP-N1) were transfected into fibroblasts using plasmid DNA (BD Biosciences Clontech, Japan) and Lipofectamine 2000 (Invitrogen, Carlsbad, CA, USA). The cultured cells were observed at 24 h, 48 h, 72 h, 96 h, 1 week, 2 weeks and 1 month after transfection. Expression of the three fluorescent protein genes was observed under a confocal microscope (Nikon TE-2000-E, Japan) with excitation wavelengths of 543 nm, 488 nm and 488 nm and 543 nm, respectively. For each group, 10 microscopic fields were considered to calculate the transfection efficiencies as the ratio of positive cells to the total cells in the field.¹³ Forty-eight h after transfection, the apoptotic rates were estimated on cell cultures after staining nuclear DNA with DAPI. The effect of exogenous gene transfection on cell viability was also assessed by the Trypan blue exclusion test.

Results

Morphological observation of Jingning chicken fibroblasts

Cells sprouted from small tissue pieces one day after being plated in the bottom of a tissue culture flask, and then continued to proliferate and were subcultured when 80-90% was confluent in about 2 days. The cells displayed typically fibrous and fusiform morphology with centrally located oval-shaped nuclei. Morphologically epithelial cells were seldom present, too. The fibroblasts grew rapidly and replaced the epithelial cells gradually after 2-3 passages, and then a relatively pure fibroblast line was obtained. The viabilities of Jingning chicken fibroblasts before freezing and after recovery were $96.23 \pm 0.58\%$ and $90.51 \pm 0.74\%$, respectively ($P < 0.5$), as estimated by the Trypan blue exclusion test (Figure 1 a,b,c).

Growth dynamics

The growth curve of Jingning chicken fibroblasts appeared sigmoidal (Figure 2) and the PDT was about 48 h. Latent phase lasted about 24 h, followed by an exponential phase 2 days after seeding, giving way to the stationary phase thereafter.

Microorganism detection

The medium remained clear for the whole

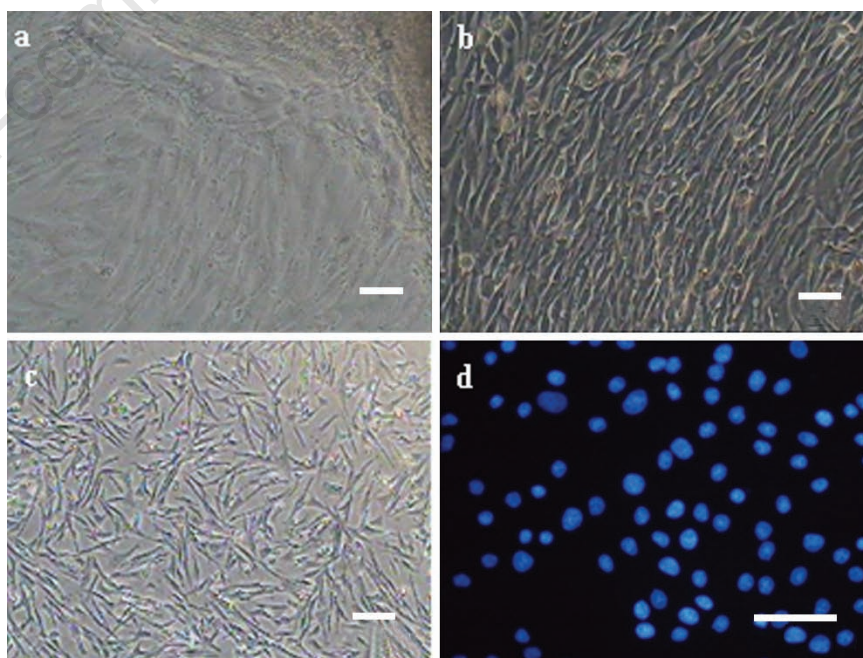


Figure 1. Morphology of Jingning chicken embryonic fibroblasts cultured *in vitro*. (a) 1-2 d after explanting; (b) before cryopreservation; (c) 24 h after recovery; (d) Jingning chicken fibroblasts stained with Hoechst 33258.

period and no abnormalities could be observed under the microscope. The results indicated that the Jingning chicken fibroblasts were free of bacterial contamination and the fibroblast cultures were apparently free of mycoplasmas. Would there be abundant punctiform and filiform blue fluorescence in the nucleoli, it could be concluded that the cells were contaminated by mycoplasmas (Figure 1d), as shown by the

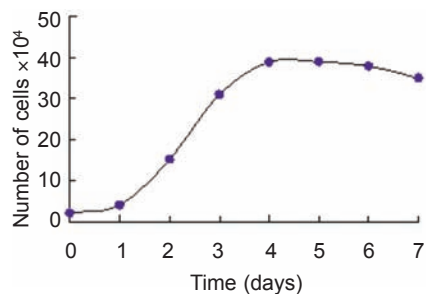


Figure 2. Growth curve of Jingning chicken embryonic fibroblasts.

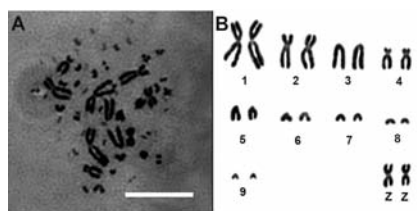


Figure 3. (A) representative spread of Jingning chicken in metaphase (♂); (B) karyotype of Jingning chicken fibroblasts (♂).

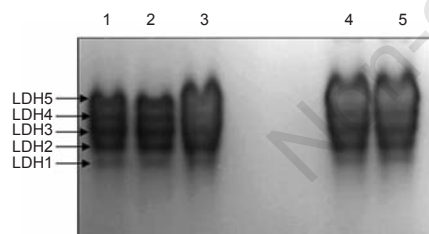


Figure 4. Isoenzyme patterns of lactate dehydrogenases from Jingning and Langshan chickens. Lanes 1 and 2, Jingning chicken; lanes 3, 4 and 5, Langshan chicken.

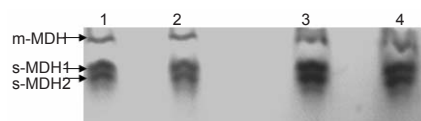


Figure 5. Isoenzyme patterns of malate dehydrogenases from Jingning and Langshan chickens. Lanes 1 and 2, Jingning chicken; lanes 3 and 4, Langshan chicken.

positive result of Li *et al.*¹⁶ The tests for virus contamination were both negative as well, as indicated by the cytopathogenic evidence and the hemadsorption test.

Karyogram and chromosome number of Jingning chicken

The chromosome number of diploid Jingning chicken is 78, including 10 pairs of macrochromosomes and 29 pairs of microchromosomes. The sex chromosome type is ZZ(♂)/ZW(♀) (Figure 3). The parameters including relative length, centromere index and kinetochore type are shown in Table 1. In this experiment, 100 representative spreads of metaphases at passages 3-5 were observed under the microscope to count the chromosome numbers, and the mean proportion of diploid cells was 97.62±2.12% (mean value ± SD).

Table 1. Chromosome parameters of Jingning chicken (♂).

Chromosome no.	Relative length (%)	Centromere index (%)	Kinetochore type
1	22.96±0.027	36.93	SM
2	17.31±0.022	37.47	SM
3	11.85±0.023	0	M
4	10.56±0.019	27.73	SM
5	6.24±0.026	0	T
6	4.91±0.035	0	T
7	4.05±0.032	0	T
8	3.22±0.024	0	T
9	2.71±0.019	0	T
Z	11.12±0.033	44.48	M
Z	11.47±0.036	44.09	M

SM, submetacentric chromosomes; T, telocentric chromosomes; M, metacentric chromosomes.

Table 2. Relative migration fonts of lactate dehydrogenase.

Breeds	LDH1 (%)	LDH2 (%)	LDH3 (%)	LDH4 (%)	LDH5 (%)
Jingning chicken	38.89	33.33	28.89	24.44	17.78
Langshan chicken	37.78	32.22	26.67	22.22	15.56

Table 3. Relative migration fonts of malate dehydrogenase.

Breeds	m-MDH (%)	s-MDH1 (%)	s-MDH2 (%)
Jingning chicken	8.1	14.52	20.97
Langshan chicken	9.68	16.13	22.58

Table 4. Transfection efficiencies of six fluorescent proteins in Jingning chicken.

Time/h	Six fluorescent genes					
	pEGFP-N3 (%)	pEGFP-C1 (%)	pEYFP-N1 (%)	pDsRed1-N1 (%)	pECFP-N1 (%)	pECFP-mito (%)
24	31.8	37.7	41.9	28.3	36.5	26.4
48	28.5	30.6	32.2	19.4	20.3	17.8
72	20.2	24.8	27.7	11.2	13.5	10.1

Isoenzyme polymorphisms

The profiles of isoenzyme polymorphisms might be characteristic in a given species or tissue.¹⁷ Polymorphism analysis of isoenzymes is currently a standard method for the quality control to characterize cell lines and prevent interspecies contamination. Isoenzyme patterns of LDH and MDH were obtained using vertical slab non-continuous PAGE, stained by Coomassie brilliant blue and compared with those from other breeds (Figures 4 and 5). The LDH RFs were in the order of LDH-5, LDH-4, LDH-3, LDH-2, LDH-1 (Figure 4); while the two bands of MDH were m-MDH and s-MDH (Figure 5). The two chicken breeds have their own characteristic bands, each one with a different migration rate (Table 2 and 3). The results indicated that there was no cross-contamination from other cell lines.

Expression of exogenous genes in Jingning chicken fibroblasts

The six fluorescent genes, pEGFP-C1, pEGFP-N3, pECFP-N1, pECFP-mito, pEYFP-N1 and pDsRed1-N1 were all highly expressed according to the optimized condition at 24 h, the transfected cells increased gradually as time passed by (Figure 6 and 7); the transfection efficiencies are listed in Table 4. Using confocal microscopy, we observed that a few positive cells began to shrink and disaggregate, and became detached and even dead. The viabilities upon transfection with pEGFP-C1, pEGFP-N3, pECFP-N1, pECFP-mito, pEYFP-N1, pDsRed1-N1 were 89.5%, 88.2%, 87.6%, 86.5%, 89.3% and 87.6%, respectively. Using the Trypan blue exclusion test, it was found that there were no significant differences ($P > 0.05$) compared with non-transfected controls (91.4%). Using DAPI, most of the cells had nuclei with evenly distributed blue fluorescence, a few only exhibiting deeply stained fragmented nuclei and apoptotic bodies (Figure 8). The apoptotic rates of cells transfected with pEGFP-N3 and pDsRed1-N1 (other apoptotic rates of cells not shown) were 1.6% and 1.5%, not significantly different ($P > 0.05$) from the control ones (1.4%).

Discussion

Chicken embryonic fibroblasts were selected in this experiment as they offer many advantages, such as high vitality and purity, easy access, low probability of bacterial contamination. A Jingning chicken fibroblast line was successfully established from 57 embryo samples by adherent culture. The biological and genetic characteristics may be altered by *in vitro* culture after many passages, so a minimal number of passages are recommended to protect the cell lines against degeneration. Morphological observation indicated that there were both epithelial cells and fibroblasts during the primary and the first several passages of the explanted tissues. Due to their different tolerance to trypsinization, the fibroblasts detached from the flasks earlier when digested with trypsin and adhered again quickly after passage, whilst most epithelial cells were difficult to adhere, or only did so in an unstable manner and fell off when vibrated.¹⁸ For this reason, a purified fibroblast line could be obtained after 2-3 passages. Isoenzyme polymorphisms and karyotyping can together effectively confirm the origin of a cell line and identify possible cross-contamination. The genetic stability of cell line is critical to preserve the genetic resources, namely the fibroblasts must maintain the same diploidy as cells *in vivo*. International standards of poultry karyotype are 8 pairs of large chromosomes, with 6 chromosomes Z and W, and 30 pairs of microchromosomes.¹⁹ Avian diploid

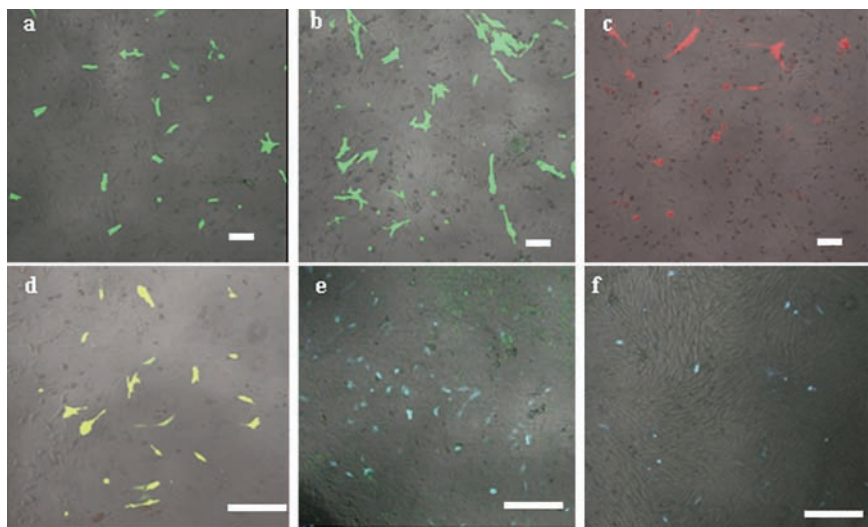


Figure 6. Comparison of six fluorescent proteins at 24 h after transfection using a confocal microscope with excitation wavelengths of 433 nm-588 nm to determine the transfection efficiencies. (a to f) were the transfection results of pEGFP-C1, pEGFP-N3, pDsRed1-N1 pEYFP-N1, pECFP-N1 and pECFP-mito respectively.

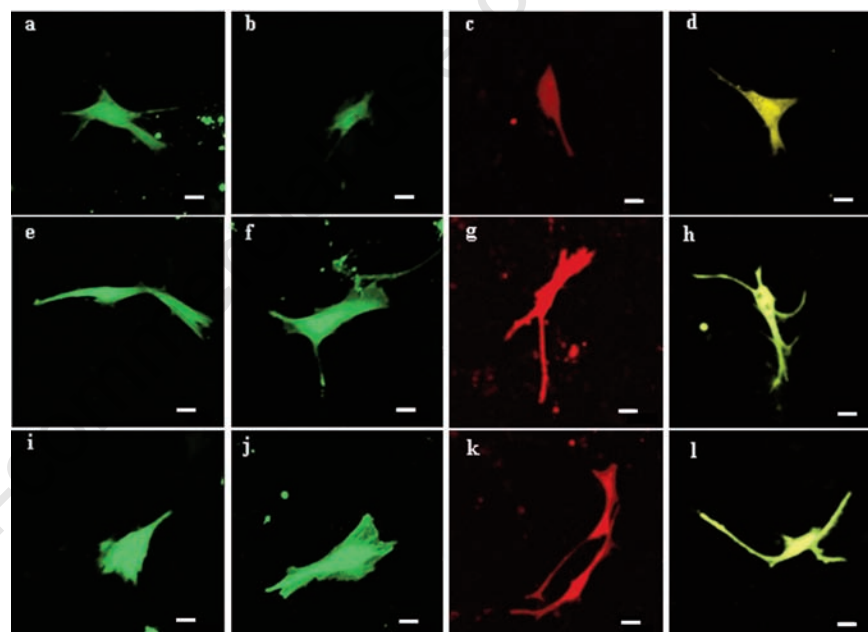


Figure 7. Expression of pEGFP-C1, pEGFP-N3, pDsRed1-N1 and pEYFP-N1 gene in Jingning chicken fibroblasts. (a-d) Fluorescent proteins of pEGFP-C1, pEGFP-N3, pDsRed1-N1 and pEYFP-N1 at 24 h after transfection; (e-h) fluorescent proteins of pEGFP-C1, pEGFP-N3, pDsRed1-N1 and pEYFP-N1 at 48 h after transfection; (i-l) fluorescent proteins of pEGFP-C1, pEGFP-N3, pDsRed1-N1 and pEYFP-N1 at 72 h after transfection.

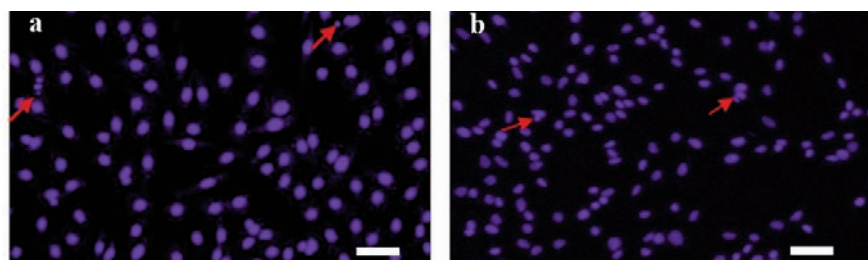


Figure 8. Apoptosis detection of pEGFP-C1 and pEYFP-N1 transfected fibroblasts stained with DAPI. Apoptosis detection of cells (a) upon pEGFP-C1 transfection and (b) upon pEYFP-N1 transfection at 24 h stained with DAPI.

chromosome number are greatly diversified, the majority of which ranged from 78 to 82. Chicken macrochromosome number is 7.8 ± 0.9 , fluctuating from 6 to 9, and the microchromosome number is 31.9 ± 2.5 , the range varying from 24 to 35. In this study, the results showed that the diploid chromosome number of Jingning chicken was $2n=78 \pm 0.9$ in 100 cells, with 10 pairs of macrochromosomes and 29 pairs of microchromosomes. The proportion of $2n=78$ cells were 97%, as detected in 100 cells. Most Jingning chicken chromosomes are tiny and may be easily lost in the preparation process or hardly identified by the interference of dye; this makes their count and even the morphological observation rather difficult. Therefore, the colchicine and hypotonic treatment should be carefully controlled in the metaphase preparation.

In cell cultures from primary explants, contamination may occur due to the undesired presence of cells from different species or tissues. Isoenzyme polymorphisms exist in different species, different races, different individuals and even different tissues of the same species. Therefore, biochemical analysis of isoenzyme polymorphisms is a standard method for the detection of cell line cross-contamination in the current important biological resource centers throughout the world, such as the ATCC, ECACC (European collection of animal cell cultures) and DSMZ (Deutsche Sammlung von Mikroorganismen und Zellkulturen).²⁰ The cell line cross-contamination can be detected when presence of other cells is higher than 10%.²¹ LDH and MDH are pivotal enzymes in glycolysis and citric acid cycle, two major metabolic pathways of animals. Genes and metabolites doubly control LDH gene expression, so the expression of LDHs had species and tissue specificity. We optimized the method of ATCC and detected the five bands of LDHs, corresponding to LDH1, LDH2, LDH3, LDH4 and LDH5. Chicken MDHs includes m-MDH and s-MDH, among which s-MDH bands are generally large, highly active and higher in content than m-MDH. In the results, MDHs of Jingning chicken embryonic fibroblast line displayed two bands, s-MDH and mMDH. The motility rate of s-MDH is higher than that of m-MDH, similar as in the early development of chicken embryos, indicating that MDH activities of *in vitro* cultured cells were similar to those of the source organs in the adults. This six enhanced fluorescent proteins in the present study are characterized with stable structure, as well as effective and germ-line independent expression. They are characterized by brighter fluorescence, more efficient expression of the transcription than lacZ, CAT and other common fluorescent markers in animal cells.²² High transfection efficiencies were obtained at 24 h. When the transfection efficiencies decreased, bright fluorescence could still be observed even after a week, indicating that the exogenous genes could be

replicated, transcribed, translated and subsequently modified in the fibroblasts. The results provided solid theoretical and technical basis for structural genomics, functional genomics and transgenic researches concerning the Jingning chicken in the future. Trypan blue tests suggested that the viabilities after transfection were non-significantly different ($P > 0.05$) from the controls (91.4%), implying that the transfection conditions used and the expression of fluorescent proteins did not affect cell growth and proliferation.

In summary, a Jingning Chicken cell bank containing biologically normal and genetically stable fibroblasts was successfully established, meeting the standards of cell line quality standards of the major international culture collection and centers. Exogenous gene expression results supported their strong vitality and rationalized their applications in transgenic therapies and genetics. The precious genetic resource of Jingning chicken was therefore well preserved at cell level, and could be used as biological materials for genetics, biomedical sciences, cell and molecular biology, immunology and so forth.

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